

# Alkaloids from Chilean species of the genus *Rhodophiala* C. Presl (Amaryllidaceae) and their chemotaxonomic importance

## Alcaloides de especies chilenas del género *Rhodophiala* C. Presl (Amaryllidaceae) y su importancia quimiotaxonómica

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### ABSTRACT

The family Amaryllidaceae is widely distributed from temperate to tropical regions. Amaryllidaceae species from the subfamily Amaryllidoideae can biosynthesize alkaloids with important physiological effects. *Rhodophiala* C. Presl is one of the native genera of Amaryllidoideae of Chile, Argentina, Paraguay, Uruguay and Brazil. However, despite the diversity of this genus in Chile, their alkaloids have only been studied previously in one species of this country. The present work aims to analyze the alkaloid profiles and chemotaxonomically compare three other Chilean species of *Rhodophiala*: *Rhodophiala bagnoldii* (Herb.) Traub, *Rhodophiala pratensis* (Poepp.) Traub and *Rhodophiala volckmannii* Phil. Bulb extracts were analyzed by means of gas chromatography-mass spectrometry (GC-MS) and alkaloids were characterized according to retention time and fragmentation pattern. The skeleton type alkaloids detected were lycorine, crinine, galanthamine, homolycorine, tazettine and montanine. All analyzed species showed different alkaloid profiles, indicating these compounds can be used as a chemotaxonomic tool. Furthermore, the alkaloid types detected in this genus have multiple reported biological properties and these species can constitute new sources of important medicinal products.

**KEYWORDS:** Alkaloids, Chilean species of Amaryllidaceae, GC-MS, *Rhodophiala*.

### RESUMEN

La familia Amaryllidaceae está ampliamente distribuida desde regiones templadas a tropicales. Las especies de la subfamilia Amaryllidoideae biosintetizan alcaloides con importantes efectos fisiológicos. *Rhodophiala* C. Presl es uno de los géneros nativos de Amaryllidoideae de Chile, Argentina, Paraguay, Uruguay y Brasil. Sin embargo, a pesar de la diversidad de este género en Chile, sólo se han estudiado los alcaloides de una especie de este país. En este trabajo se analiza el perfil alcaloideo y se comparan quimiotaxonómicamente otras tres especies chilenas del género *Rhodophiala*: *Rhodophiala bagnoldii* (Herb.) Traub, *Rhodophiala pratensis* (Poepp.) Traub y *Rhodophiala volckmannii* Phil. Se analizaron extractos de bulbos mediante cromatografía de gases-espectrometría de masas (CG-EM). Los alcaloides fueron caracterizados por su tiempo de retención y patrón de fragmentación. Los tipos de esqueletos de los alcaloides detectados fueron licorina, crinina, galantamina, homolicorina, tazetina y montanina. Todas las especies analizadas presentaron diferentes perfiles alcaloideos, por lo que estos compuestos pueden ser usados como una herramienta quimiotaxonómica. Además, los tipos de alcaloides detectados en este género tienen múltiples propiedades biológicas reportadas en literatura y estas especies pueden constituir nuevas fuentes de productos de importancia medicinal.

**PALABRAS CLAVE:** Alcaloides, especies chilenas de Amaryllidaceae, CG-EM, *Rhodophiala*.

### INTRODUCTION

The family Amaryllidaceae has three subfamilies: Amaryllidoideae, Agapanthoideae and Allioideae (Chase *et al.* 2009). Species of the subfamily Amaryllidoideae

are widely known for biosynthesis of pharmacologically active alkaloids, including antitumor, antiviral, antimarial and immunostimulant activity; as well as inhibition of the acetylcholinesterase enzyme (Bastida *et al.* 2011, Ding *et al.* 2017). These alkaloids are named Amaryllidaceae

alkaloids and classified into nine skeleton types, for which the representative alkaloids are: norbelladine, lycorine, homolycorine, crinine, haemanthamine, narciclasine, tazettine, montanine and galanthamine (Bastida *et al.* 2011).

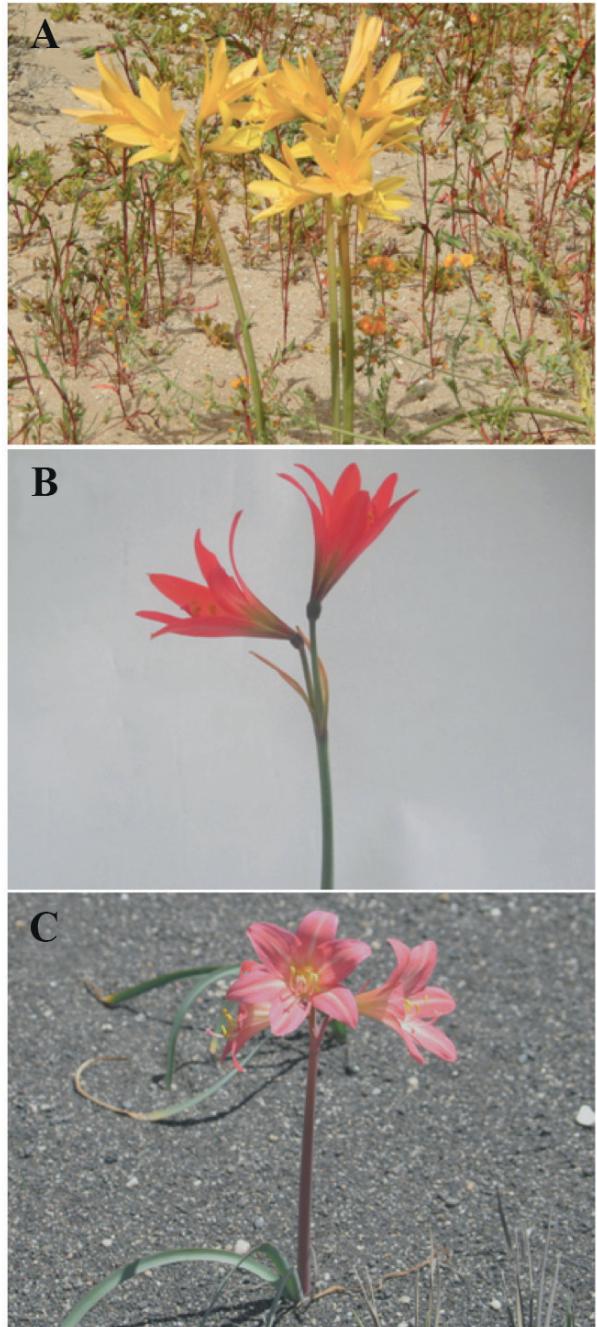


FIGURE 1. (A) *R. bagnoldii* in its natural habitat. (B) Flowers of *R. pratensis*. (C) *R. volckmannii* in its natural habitat (A and C: Photos by C. M. Baeza. B: Photo by I. Lizama-Bizama). / (A) *R. bagnoldii* en su hábitat natural. (B) Flores de *R. pratensis*. (C) *R. volckmannii* en su hábitat natural (A y C: Fotografías de C. M. Baeza. B: Fotografía de I. Lizama-Bizama).

There are more than 30 native species of the subfamily Amaryllidoideae in Chile, from Arica and Parinacota Region to Los Lagos Region (latitude 18° S to 43° S) and with a wide altitudinal distribution (Ravenna 2003, Arroyo-Leuenberger & Dutilh 2008).

*Rhodophiala* C. Presl is one of the native genera of Chile, and is also distributed in Argentina, Brazil, Paraguay and Uruguay (Arroyo-Leuenberger & Dutilh 2008). In Chile, *Rhodophiala* species are found in the coastal desert and semidesert areas, in the coastal mountain range, the central valley and the plains and hills of the foothills and mountains of the Andes, between Antofagasta Region and Los Lagos Region (Schiappacasse *et al.* 2002). This genus is characterized by a funnel-shaped perigon of 6 tepals; an androecium of 6 slightly unequal stamens, and a tricarpellary gynoecium of inferior ovary, terminating in a trifid stigma (Baeza *et al.* 2012). It also presents flowers of red, pink, orange and yellow. These plants have high ornamental value, with many lesser-known species, some of which even have conservation problems (Muñoz *et al.* 2009).

Despite the variety of Amaryllidoideae species in Chile, only two species have been chemically studied (Pacheco *et al.* 1978, 1981, 1982, Sepúlveda *et al.* 1982, Pacheco & Silva 1992). The chemistry knowledge of these species can help to solve taxonomic problems of Chilean Amaryllidoideae due to their complex morphological variation (Baeza *et al.* 2009, Fuentes *et al.* 2007). In fact, the *Rhodophiala* genus requires an urgent taxonomic revision due to doubts about the identification of several populations (Baeza *et al.* 2016).

Considering the limited knowledge on secondary metabolites regarding these native plants and the varied backgrounds on the biological activity of their alkaloids, in this work we analyze the alkaloid profiles for three species of this genus: *Rhodophiala bagnoldii* (Herb.) Traub, *Rhodophiala pratensis* (Poepp.) Traub and *Rhodophiala volckmannii* Phil. (Fig. 1). *R. bagnoldii* lives in sandy coastal and inland areas from southern Antofagasta Region to the Coquimbo Region (Schiappacasse *et al.* 2002). *R. volckmannii* is located on the coast near Cobquecura, in the Bío-Bío Region (Muñoz-Schick 1973). Meanwhile, *R. pratensis* preferably grows in sandy soils of the Andean Precordillera of the Maule and Bio-Bío Regions (Baeza *et al.* 2012). Thus, the main objective of this research was to chemotaxonically differentiate these species, contributing chemical characters to the taxonomy of this genus.

## MATERIALS AND METHODS

### PLANT MATERIAL

Plants of *Rhodophiala bagnoldii*, *R. pratensis* and *R. volckmannii* were collected. *R. bagnoldii* was collected in CHILE, Atacama Region, Huasco Province, 102 m (28°24'45"S-71°10'54"W), 19-X-2011, I. Lizama-

Bizama 9. *R. pratensis* was collected in CHILE, Bío-Bío Region, Bío-Bío Province, road to Antuco, 368 m (37°24'37"S-71°57'14"W), 27-I-2011, I. Lizama-Bizama w/n. *R. volckmannii* was collected in CHILE, Bío-Bío Region, Valle del Itata Province, Colmuyao beach, near Cobquecura, 16 m (36°09'08"S-72°48'21"W), 15-XII-2010, I. Lizama-Bizama w/n. Collections were performed during the flowering periods of these species. Specimens were deposited at the herbarium of the University of Concepción (CONC), Concepción, Chile. Bulbs were separated from the flower scapes, leaves and roots, and then minced for maceration.

#### ALKALOID EXTRACTION

Fresh bulbs (90 g of *R. pratensis*, 114 g of *R. volckmannii* and 430 g of *R. bagnoldii*) were macerated in 0.1 N HCl at room temperature for 72 h. The raw extracts were defatted three times with  $\text{CH}_2\text{Cl}_2$ . The solvent was evaporated under reduced pressure. These extracts were then basified with 25 % ammonia solution and the alkaloids extracted with  $\text{CH}_2\text{Cl}_2$ . The organic solvent was evaporated and yielded the following extracts: 37 mg of *R. pratensis*, 23 mg of *R. volckmannii* and 266 mg of *R. bagnoldii*. Each alkaloid extract was dissolved in  $\text{CHCl}_3$  to a final concentration of 10 mg/mL for further analysis.

#### GC-MS ANALYSIS

The GC-MS analysis of underivatized alkaloids was carried out on a gas chromatograph (Agilent 7890A) with a splitless injector (250 °C) and a mass detector (Agilent 5975C). An HP-5MS capillary column (30 m × 0.25 mm × 0.25  $\mu\text{m}$ ) and helium gas (constant flow 1 mL/min) were used for separation.

The temperature program was: 5 min hold at 100 °C, 100–275 °C at 13 °C/min and 32 min hold at 275 °C. Detection range was  $m/z$  50–550. The injector temperature was 250 °C. Tentative identification of the alkaloid components was accomplished by matching mass spectra to records in the NIST 05 (NIST /EPA/NIH MASS 2005 Spectral Library) and by comparing the obtained spectra with those reported in the literature. NIST 05 collects representative alkaloids of all the skeletons referenced. The structure of an alkaloid is tentatively assigned when the overlap with the database exceeds 90%. Skeletons of unidentified compounds were proposed according to the highest index of similarity with those listed in the database. Furthermore, a standard was used for galanthamine identification.

Percentage of the compounds in the extracts was calculated based on the total area of the GC-MS peaks (Berkov *et al.* 2008).

## RESULTS

A total of 25 compounds with mass spectral characteristics of Amaryllidaceae alkaloids were found in the bulbs of the studied species (Table 1). Seventeen were found in *R. bagnoldii* (Fig. 2), seven in *R. pratensis* and four in *R. volckmannii*. The retention time of alkaloids was between 18 and 24 min.

Each analyzed species showed a distinct alkaloid profile. In addition, all these species share tazettine. Only *R. bagnoldii* and *R. pratensis* share crinan-3-one. Major alkaloids for the three species were montanine type: A10 in *R. bagnoldii*, montanine in *R. pratensis* and A19 in *R. volckmannii* (Table 1).

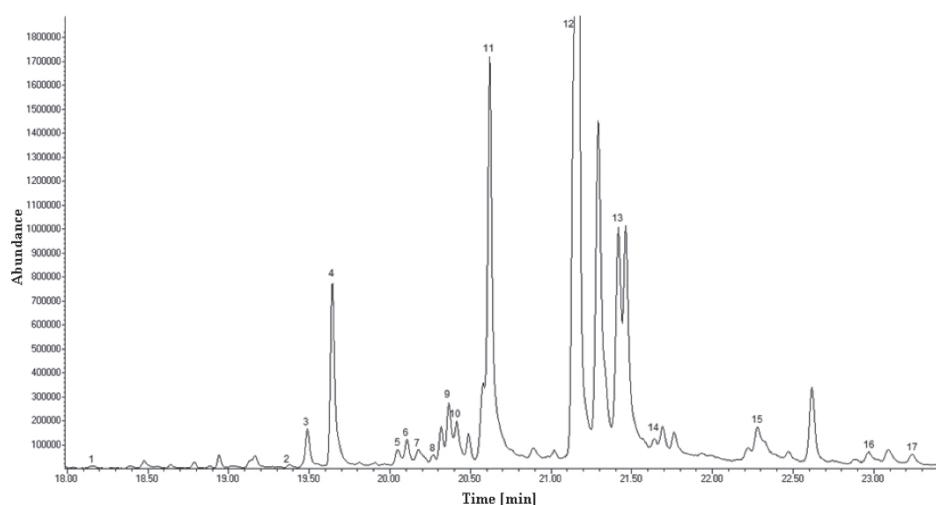


FIGURE 2. Representative chromatogram of extract for *R. bagnoldii*. 1: A1. 2: A2. 3: Galanthamine. 4: A3. 5: 1,2-didehydro-(3 $\beta$ )-Crinan-3-ol. 6: A4. 7: A5. 8: A6. 9: A7. 10: A8. 11: A9. 12: A10. 13: Tazettine. 14: A11. 15: Lycorine. 16: A12. 17: Crinan-3-one. / Cromatograma representativo del extracto de *R. bagnoldii*. 1: A1. 2: A2. 3: Galantamina. 4: A3. 5: 1,2-didehidro-(3 $\beta$ )-Crinan-3-ol. 6: A4. 7: A5. 8: A6. 9: A7. 10: A8. 11: A9. 12: A10. 13: Tazetina. 14: A11. 15: Licorina. 16: A12. 17: Crinan-3-ona.

TABLE 1. Alkaloids in species of the genus *Rhodophiala*. / Alcaloides en especies del género *Rhodophiala*.

RT <sup>a</sup>	ALKALOID (SKELETON TYPE)	m/z (% RELATIVE INTENSITY)	% <sup>b</sup>
<i>Rhodophiala bagnoldii</i> :			
18.16	A1 (Lycorine)	269 (56), 268 (100), 241 (17), 224 (23), 209 (14), 207 (21), 196 (15), 182 (16), 167 (19), 84 (15)	0.09
19.38	A2 (Montanine)	315 (87), 285 (53), 284 (55), 281 (33), 268 (45), 207 (63), 134 (100), 132 (55), 96 (36), 70 (40)	0.10
19.49	Galanthamine * (Galanthamine)	288 (15), 287 (79), 286 (100), 270 (16), 244 (27), 230 (12), 216 (38), 174 (33), 128 (11), 115 (16)	1.17
19.65	A3 (Galanthamine)	290 (10), 289 (59), 288 (100), 232 (7), 213 (6), 202 (10), 189 (7), 188 (8), 187 (9), 115 (10)	5.05
20.05	1,2-didehydro-(3β)- Crinan-3-ol (Crinine)	271 (100), 254 (25), 252 (27), 228 (28), 200 (26), 199 (80), 187 (65), 173 (25), 128 (27), 115 (29)	0.47
20.10	A4 (Homolycorine)	285 (2), 284 (2), 207 (5), 115 (2), 110 (9), 109 (100), 108 (17), 94 (4), 93 (2), 77 (2)	0.60
20.18	A5 (Tazettine)	315 (20), 300 (44), 232 (25), 231 (100), 230 (19), 207 (18), 185 (17), 115 (21), 109 (59), 70 (68)	0.48
20.27	A6 (Crinine)	303 (100), 302 (26), 272 (46), 242 (22), 218 (37), 217 (38), 216 (20), 207 (23), 202 (21), 201 (30)	0.15
20.36	A7 (Crinine)	288 (23), 287 (97), 244 (35), 217 (20), 216 (27), 215 (100), 203 (58), 189 (22), 128 (22), 115 (25)	1.57
20.41	A8 (Montanine)	301 (100), 271 (38), 254 (53), 252 (33), 212 (31), 199 (43), 185 (68), 141 (35), 128 (31), 115 (38)	1.06
20.62	A9 (Crinine)	290 (19), 289 (100), 288 (27), 272 (21), 260 (23), 218 (47), 217 (35), 216 (18), 202 (23), 201 (24)	14.48
21.17	A10 (Montanine)	319 (23), 301 (100), 270 (88), 257 (36), 229 (24), 226 (28), 223 (26), 199 (21), 185 (31), 115 (22)	27.23
21.42	Tazettine (Tazettine)	331 (24), 298 (18), 248 (16), 247 (100), 201 (16), 199 (15), 181 (13), 115 (15), 71 (21), 70 (23)	7.27
21.64	A11 (Lycorine)	305 (16), 301 (15), 300 (14), 252 (14), 250 (16), 228 (18), 227 (78), 226 (100), 207 (22), 147 (17)	0.47
22.28	Lycorine (Lycorine)	287 (19), 286 (16), 268 (20), 250 (16), 242 (11), 228 (13), 227 (63), 226 (100), 207 (11), 147 (11)	1.77
22.97	A12 (Homolycorine)	281 (3), 207 (7), 193 (2), 165 (2), 110 (10), 109 (100), 108 (18), 94 (2), 82 (3), 81 (2)	0.48
23.24	Crinan-3-one (Crinine)	284 (25), 281 (17), 271 (100), 270 (55), 240 (20), 211 (21), 207 (37), 181 (57), 153 (17), 152 (21)	0.39
<i>Rhodophiala pratensis</i> :			
19.99	A13 (Homolycorine)	135 (5), 117 (7), 110 (10), 109 (100), 108 (16), 81 (5), 71 (4), 67 (4), 57 (8), 55 (7)	1.06
20.06	A14 (Tazettine)	315 (18), 300 (38), 231 (100), 135 (27), 115 (25), 109 (27), 91 (19), 70 (79), 57 (19), 55 (27)	0.57
20.23	A15 (Crinine)	269 (100), 240 (33), 212 (27), 186 (91), 185 (45), 135 (29), 128 (71), 127 (31), 115 (34), 55 (35)	0.51
20.79	A16 (Crinine)	295 (24), 271 (100), 270 (44), 211 (24), 181 (85), 153 (27), 152 (31), 135 (26), 94 (38), 55 (25)	0.66
21.07	Montanine (Montanine)	302 (23), 301 (100), 270 (91), 257 (36), 229 (26), 226 (31), 223 (29), 185 (36), 128 (24), 115 (28)	5.90
21.29	Tazettine (Tazettine)	331 (22), 298 (17), 248 (16), 247 (100), 201 (18), 199 (16), 181 (16), 115 (23), 71 (26), 70 (29)	2.07
23.06	Crinan-3-one (Crinine)	272 (21), 271 (100), 270 (44), 207 (29), 181 (65), 135 (29), 91 (25), 81 (21), 67 (24), 55 (30)	0.30

RT <sup>a</sup>	ALKALOID (SKELETON TYPE)	m/z (% RELATIVE INTENSITY)	% <sup>b</sup>
<i>Rhodophiala volckmannii</i> :			
18.08	A17 (Lycorine)	269 (55), 268 (100), 135 (20), 91 (22), 83 (21), 77 (16), 69 (19), 67 (17), 57 (17), 55 (28)	0.56
20.23	A18 (Crinine)	269 (92), 186 (91), 135 (78), 128 (68), 91 (59), 83 (58), 81 (64), 69 (69), 57 (56), 55 (100)	0.28
21.08	A19 (Montanine)	301 (100), 270 (94), 257 (36), 223 (40), 207 (36), 185 (47), 135 (46), 115 (43), 77 (37), 55 (56)	2.02
21.29	Tazettine (Tazettine)	331 (25), 298 (22), 248 (21), 247 (100), 135 (23), 115 (30), 71 (33), 70 (33), 57 (24), 55 (26)	1.08

<sup>a</sup>Retention time (min). / Tiempo de retención (min).

<sup>b</sup>Percentage of the compound in the extract. Values are expressed as a percentage of the total ion chromatogram. / Porcentaje del compuesto en el extracto. Los valores están expresados como un porcentaje del cromatograma total de iones.

\*Comparison with a standard MS spectra. / Comparación con un espectro EM estándar.

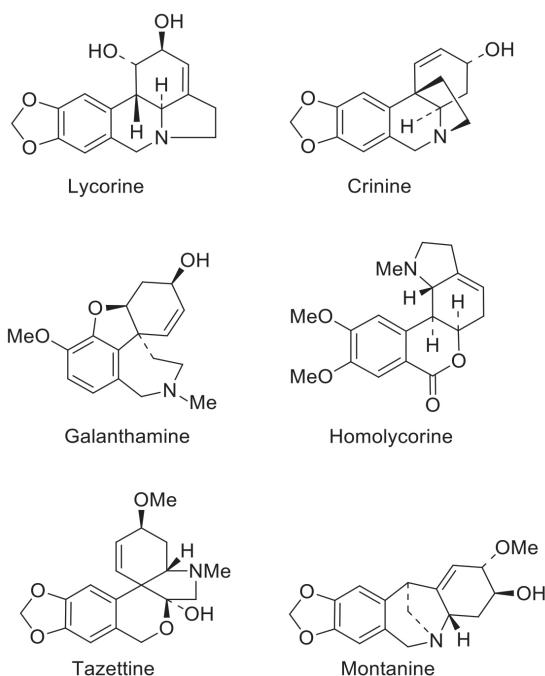


FIGURE 3. Types of Amaryllidaceae alkaloids identified in the extracts: lycorine, crinine, galanthamine, homolycorine, tazettine and montanine. / Tipos de alcaloides de Amaryllidaceae identificados en los extractos: licorina, crinina, galantamina, homolicorina, tazetina y montanina.

The skeleton type alkaloids detected were tentatively identified as lycorine, crinine, galanthamine, homolycorine, tazettine and montanine (Fig. 3, Table 2). *R. bagnoldii* bulbs had the highest variety of skeleton types. Galanthamine-type alkaloids were present only in *R. bagnoldii*. *R. pratensis* presented crinine-type and homolycorine-type alkaloids, in addition to tazettine-type and montanine-type alkaloids. Meanwhile, only *R. volckmannii* and *R. bagnoldii* presented lycorine-type alkaloids.

TABLE 2. Presence of different types of Amaryllidaceae alkaloids in the studied species. / Presencia de los diferentes tipos de alcaloides de Amaryllidaceae en las especies estudiadas.

SKELETON TYPE	<i>R. BAGNOLDII</i>	<i>R. PRATENSIS</i>	<i>R. VOLCKMANNII</i>
CRININE	✓	✓	✓
GALANTHAMINE	✓	-	-
HOMOLYCORINE	✓	✓	-
LYCORINE	✓	-	✓
MONTANINE	✓	✓	✓
TAZETTINE	✓	✓	✓

## DISCUSSION

Three species of *Rhodophiala*, *R. bagnoldii*, *R. pratensis* and *R. volckmannii*, share alkaloids with crinine, tazettine and montanine skeleton types (Table 2). The presence of these three skeleton types could be characteristic of the *Rhodophiala* genus, in comparison to other genera of Chilean Amaryllidoideae. Thus, the Amaryllidaceae alkaloids could allow to differentiate Chilean Amarylloideae genera. On the other hand, only *R. bagnoldii* presented galanthamine-type alkaloids. The exclusive galanthamine-type alkaloids detected in *R. bagnoldii* could allow it to be chemotaxonically differentiated. Lycorine-type alkaloids were absent only in *R. pratensis*, and homolycorine-type alkaloids were absent only in *R. volckmannii*. The

absence and presence of these alkaloid types can be used as a chemotaxonomic tool for differentiating *Rhodophiala* species.

*R. bagnoldii* presents more morphological differences compared to the other two species studied and has a more distant geographical distribution in northern Chile. This species presents flowers with infundibuliform perigonum, of pure yellow color, or with orange lines and the greenish base of the tube (Schiappacasse *et al.* 2002). In contrast, *R. pratensis* and *R. volckmannii* are both located in the Bío-Bío Region of central Chile and have similar morphological characters. *R. pratensis* presents red scarlet flowers, sometimes yellowish (Baeza *et al.* 2012), and the studied population of *R. volckmannii* presents pale red and more bell-shaped flowers. The morphological differences and similarities between these three species are concordant with the characteristics of their alkaloid profiles: *R. pratensis* and *R. volckmannii* present greater similarity in their types of alkaloids, in comparison with *R. bagnoldii*. It is necessary to analyze more species of this genus to corroborate if they also differ in their types of Amaryllidaceae alkaloids. If there were no major differences between the types of alkaloids of all Chilean *Rhodophiala* species, these compounds could also have chemotaxonomic importance at the genus level, and thus contribute to the resolution of the taxonomic problem of the Chilean Amaryllidoideae.

According to previous studies, lycorine-type, homolycorine-type and haemanthamine-type alkaloids were found in the Chilean species *Rhodophiala ananuca* (= *Hippeastrum ananuca*) (Pacheco *et al.* 1982). In our study, only haemanthamine-type alkaloids were not found in the three analyzed species. Crinine-type, tazettine-type and montanine-type alkaloids shared by *R. bagnoldii*, *R. pratensis* and *R. volckmannii*, were not detected in the species studied by Pacheco *et al.* 1982. Our study is the first to analyze alkaloids of Chilean *Rhodophiala* by GC-MS. This method is efficient and reliable for studies on Amaryllidaceae alkaloids (Berkov *et al.* 2008).

Furthermore, it is important to mention that the alkaloid types identified in this genus have multiple and documented biological properties, and therefore these species can constitute new sources for products of medicinal importance.

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